Tandem Silica Gel-Catalysed Rearrangements and Subsequent Baeyer–Villiger **Reactions of Artemisinin Derivatives**

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Novel silica gel-catalysed reactions of dihydroartemisinin 1b to deoxyartemisinin 11 and 11β-hydroxy-11-epidihydroartemisinin 8 to compound 9 under mild conditions are described. The structures of the products were determined by mass spectrometry and 1D- and 2D-NMR spectroscopy. A mechanism for their formation is proposed.

Isolation, by Chinese investigators, of artemisinin 1a (Fig. 1) and their demonstration that it was effective in treating patients infected with drug-resistant strains of Plasmodium falciparum,

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Fig. 1 Artemisinin 1a and related compounds

even those with cerebral malaria, prompted structure-activity investigations of artemisinin derivatives.¹ These studies established that the labile peroxide moiety was required for antimalarial activity.^{1a,b} Many derivatives^{1b} of dihydroartemisinin 1b were prepared by converting the free hydroxy group into a variety of ethers, esters, carbonates, urethanes, etc. In searching for new derivatives several investigators prepared anhydrodihydroartemisinin 2 from compound 1b and attempted to convert the double bond into an epoxide 3. When Lin et al.^{2a} treated compound 2 with m-chloroperbenzoic acid (MCPBA) they found that the epoxides reacted with m-chlorobenzoic acid, formed in the reaction, to yield a mixture of two hydroxy esters. Later, Petrov and Ognyanov^{2b} successfully isolated the 11β,-12 β -epoxide 3 by reaction of compound 2 with a complex of MCPBA and KF.

In order to prepare 11β -hydroxy-11-epidihydroartemisinin 8, an aqueous acetone solution of epoxide 3 was treated with dil. aq. sulfuric acid. The resulting diol, however, proved surprisingly difficult to purify by silica gel chromatography. During chromatography diol 8 underwent an unusual silica gelcatalysed rearrangement and Baeyer-Villiger reaction to form a less polar compound 9. The structure of compound 9 has been determined and a mechanism for its formation is suggested. The reactions of dihydroartemisinin 1b and 11a-hydroxydihydroartemisinin 10 with silica gel under similar conditions were investigated and the structure of a product obtained from compound 1b was identified.

Results and discussion

Treatment of crude epoxide 3, prepared as described by Petrov and Ognyanov,^{2b} with ethanol and toluene-p-sulfonic acid yielded a mixture of hydroxy ethers, which were separated by chromatography. Analysis of their ¹H NMR spectra indicated they were epimeric at C-11, analogous to the isomeric 11-hydroxy-12B-O-(m-chlorobenzoyl)dihydroartemisinins prepared by Lin et al.^{2a} These observations suggested that both the 11α , 12α - and the 11β , 12β -epoxide were present in the crude reaction mixture. A careful examination of the ¹H NMR spectrum of the crude reaction mixture revealed that it contained 20–25% of the 11α , 12α -epoxide. In attempting to separate the mixture of epoxides by chromatography on silica gel, we found that the α -epoxide reacted with water more rapidly than did the β -isomer. A pure sample of β -epoxide 3 was obtained from the crude epoxide mixture by flash chromatography over silica gel. During flash chromatography, the 11α , 12α epoxide was selectively converted into 11a-hydroxydihydroartemisinin 10, which was eluted after epoxide 3 with more polar solvent mixtures.

Treatment of epoxide 3 with aq. acetone containing a trace of sulfuric acid yielded 11β-hydroxy-11-epidihydroartemisinin 8, which was purified by flash chromatography. If diol 8 remained in contact with silica gel it was slowly converted into the rearranged product 9. Investigation of the conditions that convert diol 8 into compound 9 revealed that the reaction was catalysed by silica gel in propan-1-ol, chloroform or (most rapidly) benzene. A 70% yield of compound 9 was obtained by refluxing a benzene solution of diol 8 with silica gel for 10 min.

Molecular-ion measurements by EI and CI mass spectrometry with a variety of gases, showed diol 8 lost CH₂O₂ in its conversion into compound 9. The latter's structure and stereochemistry were deduced from the following spectroscopic data. The ¹H NMR spectrum showed the presence of three methyl signals: a doublet at δ 0.96 (10-Me) and singlets at 1.35 (11-Me) and 1.58 (15-Me), and a single proton at 5.65 (5-H). There was no signal observed for 12-H. The ¹³C NMR spectrum of compound 9 showed 14 carbons, *i.e.* the loss of one carbon signal in agreement with mass spectral data. Tentative assignments of the ¹³C resonances in compound 9 were made (see Table 1) from distortionless enhancement by polarisation transfer (DEPT) and 2D heteronuclear multiple quantum coherence (HMQC) spectra. The assignments were verified by an analysis of the 2D heteronuclear multiple bond coherence (HMBC) spectrum (Fig. 2).

The absence, in the ¹³C NMR spectrum, of the C-12 resonance in compound 9 and that of 12-H in the ¹H NMR





Scheme 1 Proposed mechanism for silica gel-catalysed transformation of the artemisinin derivatives and subsequent Baeyer-Villiger reactions

Table 1¹³C NMR Spectral data and assignments of compounds 8(8a + 8b), 10 (10a + 10b) and 9

	8 ^{<i>a</i>}		10 <i>ª</i>		
Carbon	8a	8b	10a	10b	9
1	52.24	52.02	52.29	51.37	42.19
2	24.50	24.42	25.47	25.05	23.37
3	36.49	36.33	36.36	36.24	34.59
4	103.5	104.4	104.3	104.5	111.2
5	88.57	91.66	87.35	90.82	103.4
6	81.80	82.00	83.44	83.01	94.18
7	50.54	50.62	49.17	49.47	49.93
8	23.31	22.96	24.51	24.51	26.25
9	34.08	33.97	34.37	33.95	32.27
10	37.42	37.42	37.38	37.38	35.36
11	70.74	71.30	70.97	71.85	110.7
12	96.09	93.66	100.1	93.81	
13	28.12	29.66	22.66	22.05	21.88
14	20.05	20.19	20.34	20.23	18.64
15	25.68	25.53	25.89	25.89	24.66

"The special assignments were made from a mixture of diastereoisomeric hemiacetals in equilibrium.

spectrum suggested that the CH_2O group at C-12 was lost during the reaction. Removal of C-12 and the atoms attached to it requires the oxygen previously bonded to C-5 and C-12 to become bonded to C-11, forming a tetrahydrofuran. This deduction is consistent with the 40 ppm downfield shift of C-11 in the ¹³C NMR spectrum of compound 9 compared with that in compound 8. The loss of a second oxygen atom has been accounted for by transforming the 4,6-peroxide into a 4,6-oxide, a reaction reported by Posner and Oh for their synthetic 1,2,4trioxane.³ Support for the presence of a 4,6-oxide came from a comparison of the proton and carbon chemical shifts for 15-Me,



Fig. 2 Selected H-C couplings in the HMBC spectrum of compound 9

C-4 and C-6 in several peroxides and the corresponding oxides (Table 2). The transformation of the peroxide into an oxide in these artemisinin derivatives is usually accompanied by a 2–4 ppm downfield shift of C-4 and C-6 in the ¹³C NMR spectrum and a 0.1 ppm shift for 15-Me. The appropriate shifts in compound 9 are seen for C-4 and C-15 but the larger shift for C-6 may result from contraction of the D-ring as seen in Table 1. The stereochemistry for C-13 in compound 9 was initially assigned based on a comparison of its ¹³C NMR chemical shift with that for the methyl groups in 11β-hydroxy- and 11α-hydroxydihydroartemisinin. The ¹³C NMR chemical shifts of α-methyl groups vary from δ_C 28 to 30 whereas those for β -methyls vary between δ_C 22 and 23. Similar shifts were

 Table 2
 Selected NMR spectral data of the 4,6-peroxides and 4,6-oxides

Compound	Me at C-4 ($\delta_{\rm H}$)	C-4 ($\delta_{\rm C}$)	C-6 (δ_c)
1a	1.44	105.3	79.5
11	1.53	109.2	82.5
1b	1.43	104.1	80.3
6	1.54	108.1	82.4
Arteether 7	1.42	104.0	81.2
Deoxyarteether	1.54	107.9	83.4



No reaction Scheme 2 Silica gel-catalysed reactions of compounds 8c and 10c. (a) A 1,3-cis relationship between 6-OOH and 11-CHO favours addition of the hydroperoxide to the aldehyde. (b) A 1,3-trans relationship between 6-OOH and 11-CHO does not permit addition of the hydroperoxide to

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observed by Lin *et al.* in the reaction products of compound **2** with MCPBA.^{2a} The observed chemical shift (δ_C 22) for C-13 in compound **9** is closer to that of a β -methyl than to that for an α -methyl. The same stereochemical assignment can be deduced for compound **9** from data on the ¹H NMR chemical shift of the hydroxy group (δ 5.2) on C-11 compared with δ 2.3 for diol **8** and δ 4.6 in diol **10**. The 11 α -configuration of the hydroxy group was verified by a 2D NOESY spectrum which showed a cross-peak between the hydroxy group and 7-H. The differences

in the 13 C NMR shifts for C-5, C-6 and C-7 in compound 9 compared with diol 8 presumably are a consequence of the D-ring contracting from a six- to five-membered ring.

The HMBC spectrum was employed for unequivocal assignments of the 13 C NMR spectrum of compound 9 as seen in Fig. 2. The two-bond coupling of 5-H to C-6 identified C-6, and the three-bond coupling to C-11 requires the presence of an oxygen atom between C-5 and C-11. The stereochemical assignment of C-13 follows from that for the hydroxy group discussed above. The above spectra showed no features suggesting skeletal rearrangements.

Additional information on the reaction was obtained from unsuccessful attempts to catalyse the conversion of diol 8 into alcohol 9 by radical initiators. Heating of a chloroform solution of diol 8 in the presence of azoisobutyronitrile (AIBN) did not generate compound 9. In an effort to effect analogous transformation in artemisinin derivatives, benzene solutions of compounds 1a, 1b, arteether 7 and compound 10 were refluxed in the presence of silica gel. Artemisinin 1a, arteether 7 and compound 10 were recovered unchanged after being heated for up to 10 h. However, dihydroartemisinin 1b yielded a less polar compound, 11, in 29% yield. A molecular mass determination of compound 11 showed it had formed from substrate 1b by loss of the elements of water. An examination of its ¹H and ¹³C NMR spectra enabled us to identify compound 11 as deoxyartemisinin.⁴ This compound has been isolated from Artemisa annua⁵ and as a metabolite from the urine of malaria patients treated with artemisinin.6

A literature search for silica gel-catalysed reactions of artemisinin derivatives revealed a paper by El-Feraly et al.⁷ describing the rearrangement of dihydroartemisitene 4 to the aldehyde 5, whose structure was determined by X-ray crystallography. On the basis of our findings and El-Feraly's report, formation of compounds 9 and 11, as well as of those compounds described in the literature, have been rationalized (Scheme 1). Under neutral and/or weakly acidic conditions the hemiacetal at C-12 is in equilibrium with structures (a), (b) and (c) as shown in Scheme 1. Compound 4 undergoes the reaction sequence 4(a) to 4(b) to 4(c) followed by addition of the hydroperoxide to the C-12 aldehyde to form 4(d), e.g. compound 5. The reaction scheme thus accounts for El-Feraly's observations. Structures such as 5 have been proposed as intermediates in Baever-Villiger reactions.⁸ An intermediate such as 8(d) would be expected to undergo a Baeyer-Villiger reaction to form 8(e), which then loses formic acid to form 8(f) and then hemiacetal 9. The failure of peroxide 5 to undergo a Baeyer-Villiger reaction might be due to the presence of the adjacent carbon-carbon double bond or to a difference in the reaction conditions used by El-Feraly et al.⁷ To account for the observation that diol 10(c) does not undergo an analogous transformation, we suggest that perhaps in the acetal 10(i) (Scheme 2) the distance between the hydroperoxide and the aldehyde group is too large for formation of the key Baeyer-Villiger reaction intermediate.

The transformation of 1b to lactone 11 was rationalized by assuming that the Baeyer-Villiger reaction proceeded by a hydride migration to form compound 1b(g). Under the acidic conditions used, the hydroxy group of the hemiacetal 1b(h) and the carboxy group react to form lactone 11.

Posner and Oh suggested that the biotransformations of substrates 1a or 1b to 4,6-oxide 6 occur by radical reactions.³ However, it is also possible to account for these transformations by non-radical processes as shown in Scheme 1 for the conversion of compound 1b into lactone 11. Transformation of dihydroartemisinin 1b that is catalysed by weak acids on the surface of silica gel might also occur on cell surfaces, leading to loss of the peroxide group and as a consequence to the loss of the antimalarial activity of these drugs.

Experimental

Unless otherwise stated, the m.p.s were determined using a Reichert hot-stage apparatus and are uncorrected. Sodium borohydride, phosphorus pentaoxide, potassium fluoride and MCPBA (80-90%) were purchased from Aldrich and used without purification. All other reagents were ACS grade or the highest quality material available. Silica gel TLC plates and TLC-grade silica gel (cat. no. 10050) were from Analtech, Newark, DE 19713. Dichloromethane was dried over P_2O_5 and distilled. ¹H NMR and ¹³C NMR spectra were run on a VXR-500 or Gemini 300 spectrometer using Me₄Si as a standard in CDCl₃. J-Values are given in Hz. FT-IR spectra were measured on a Bio-Rad FTS-45 spectrometer, and CI-MS were determined on a Finnagan 4600 Mass Spectrometer. Microanalyses were performed by Galbraith Laboratory, Inc., Tennessee. Optical solutions (reported in units of 10^{-1} deg cm² g⁻¹) were measured on a Perkin Elmer 241 MC polarimeter.

MCPBA·2KF Complex.—A mixture of finely grounded potassium fluoride (2.2 g, 0.038 mol) and MCPBA (3.2 g, 0.016 mol) were suspended in CH_2Cl_2 (110 cm³), and stirred at room temperature for 7 h. The precipitate (MCPBA·2KF) was filtered off, washed with dry CH_2Cl_2 , and stored at -20 °C until used.

Epoxide 3 and 11α -Hydroxydihydroartemisinin 10.—The MCPBA·2KF complex (3.0 g) and compound 2^{2b} (600 mg, 2.25 mmol) were suspended in dry CH_2Cl_2 (70 cm³). The suspension was stirred at 4 °C for 66 h, then was filtered through a layer of anhydrous NaHCO₃. The filtrate was evaporated to dryness (570 mg) and epoxide 3 was purified by flash chromatography on a Buchner funnel (300 cm³) tightly packed with TLC-grade silica gel with hexane-ethyl acetate (5:1) as eluent. Compound 3 [(460 mg, 70%), m.p. 117-119 °C (lit., ^{2b} 117-120 °C)] was eluted first, and further washing with hexane-acetone (7:3) afforded compound 10 (115 mg, 17%), m.p. 145–150 °C (Found: C, 60.25; H, 8.1. $C_{15}H_{24}O_6$ requires C, 59.98; H, 8.05%); $[\alpha]_D + 82(c 0.22, c 0.22)$ CHCl₃); CI-MS (NH₃): 318 (M + NH₄⁺, 100%) and 300 $(M + NH_4^+ - H_2O, 75); \delta_H$ 10a: 5.40 (1 H, s, 5-H), 4.76 (1 H, s, 12-H), 4.36 and 3.86 (2 H, s, D₂O-exchangeable, 11- and 12-OH), 2.4-0.9 (11 H, overlapping carbon-skeleton protons), 1.46 (3 H, s, 15-H), 1.12 (3 H, s, 13-H) and 0.97 (3 H, d, J 5.8, 14-H₃); 10b: 5.69 (1 H, s, 5-H), 5.12 (1 H, s, 12-H), 4.68 and 3.0 (2 H, s, D₂O-exchangeable, 11- and 12-OH), 2.4-0.9 (11 H, overlapping carbon-skeleton protons), 1.46 (3 H, s, 15-H), 1.21 (3 H, s, 13-H) and 0.96 (3 H, d, J 5.8, 14-H); $\delta_{\rm C}$ (Table 1).

11β-Hydroxy-11-epidihydroartemisinin 8.—To a solution of epoxide 3 (500 mg, 1.77 mmol) in acetone (120 cm³)-water (40 cm³) was added 1 mol dm⁻³ H₂SO₄ (0.1 cm³). The solution was stirred at room temperature for 5 min and was then extracted with CH₂Cl₂. The extract was washed with 1% aq. NaHCO₃, dried over Na₂SO₄, and evaporated to dryness. Flash chromatography on silica gel (TLC grade) with hexaneacetone (3:1) as eluent yielded diol 8 (430 mg, 80%), m.p. 133-135 °C (Found: C, 60.1; H, 8.1. C₁₅H₂₄O₆ requires C, 59.98; H, 8.05%); $[\alpha]_D$ +70 (c 0.30, CHCl₃); CI-MS (NH₃): 318 (M + NH₄⁺, 40%) and 300 (M + NH₄⁺ - H₂O, 60); δ_H 8a: 5.37 (1 H, s, 5-H), 4.95 (1 H, s, 12-H), 3.35 and 2.30 (2 H, s, D₂Oexchangeable, 12- and 11-OH), 2.4–0.9 (11 H, overlapping carbon-skeleton protons), 1.53 (3 H, s, 13-H), 1.40 (3 H, s, 15-H) and 0.94 (3 H, d, J 5.8, 14-H); 8b: 5.51 (1 H, s, 5-H), 5.26 (1 H, s, 12-H), 3.80 and 2.30 (2 H, s, D₂O-exchangeable, 12- and 11-OH), 2.4–0.9 (11 H, overlapping carbon-skeleton protons), 1.45 (3 H, s, 13-H), 1.40 (3 H, s, 15-H) and 0.94 (3 H, d, J 5.8, 14-H); $\delta_{\rm C}$ (Table 1).

Preparation of Oxide 9.—To a solution of diol 8 (50.0 mg, 0.17 mmol) in benzene (5.0 cm³) was added TLC-grade silica gel (500 mg). The mixture was reluxed for 10 min, then filtered, and the silica gel was washed with acetone. The filtrate and washings were combined and evaporated to dryness. Purification on a preparative TLC (PLC) plate with hexane-ethyl acetate (7:3) as developer gave compound 9 (20.0 mg, 70% based on diol 8 consumed in the reaction) as an oil (Found: M⁺, 254.1525. C₁₄H₂₂O₄ requires M, 254.1517); [α]_D -65 (*c* 0.85, CHCl₃); δ _H 5.65 (1 H, s, 5-H), 5.20 (1 H, s, D₂O-exchangeable, 11-OH), 2.3-1.0 (11 H, overlapping carbon-skeleton protons), 1.58 (3 H, s, 15-H), 1.35 (3 H, s, 13-H) and 0.96 (3 H, d, J 5.5, 14-H); δ _C (Table 1).

Preparation of Lactone 11 from Hemiacetal 1b.—To a solution of compound 1b (50.0 mg, 0.18 mmol) in benzene (5.0 cm³) was added TLC-grade silica gel (500 mg). The mixture was refluxed for 6 h, then filtered, and the silica gel was washed with acetone–methanol (5:1). The filtrate and washings were combined and evaporated to dryness. Purification on a PLC plate with hexane–ethyl acetate (7:3) as developer afforded lactone 11 (12.0 mg, 29% based on the consumed starting material) and recovered compound 1b (8.0 mg). Compound 11: m.p. 110–112 °C (lit.,^{1e} 111–113 °C); $\delta_{\rm H}$ 5.70 (1 H, s, 5-H), 3.20 (1 H, m, 11-H), 1.53 (3 H, s, 15-H), 1.20 (3 H, d, J 7.2, 13-H) and 0.94 (3 H, d, J 5.6, 14-H).

Note added in proof: J. K. Baker, J. D. McChesney and H. T. Chi (*Pharm. Res.*, 1993, **10**, 662) have reported the isolation of **1d** (Scheme 1) by treating arteether with HCl (2.5 mol dm^{-1}) in ethanol-water (1:1).

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